

## Abstract

Effects of hydrodynamic stress, dissolved oxygen (DO) concentration and carbon sources on heterotrophic  $\alpha$ -tocopherol production by *Euglena gracilis* were investigated. In a jar fermentor without baffle plates, increasing the agitation speed up to 500 rpm had no significant effect on cell growth and  $\alpha$ -tocopherol production. However, in a jar fermentor equipped with baffle plates, both the cell growth and  $\alpha$ -tocopherol production were highly suppressed at 500 rpm. At high hydrodynamic stress, the cells secreted nucleic acid-related substances to the culture broth and the shape of the cells shifted from elongated toward spherical. High DO concentration had adverse effects on both cell growth and  $\alpha$ -tocopherol production, the optimum DO concentration being below 0.8 ppm. In comparison with glucose, the growth rate was lower but the  $\alpha$ -tocopherol content of the cells was almost four times higher when ethanol was used as the organic carbon source. In a fed-batch culture with ethanol, a very high cell concentration of 39.5 g L<sup>-1</sup> was obtained with  $\alpha$ -tocopherol content of 1200  $\mu$ g g-cell<sup>-1</sup>. This  $\alpha$ -tocopherol content is very close to the values reported for photoautotrophic and photoheterotrophic cultures. A very high  $\alpha$ -tocopherol productivity of 102  $\mu$ g L<sup>-1</sup> h<sup>-1</sup> was obtained, indicating that heterotrophic cultivation of *E. gracilis* has a very high potential as a substitute for the current method of extraction from vegetable oils.

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